Making Formalin-Fixed, Paraffin Embedded Blocks  
<https://link.springer.com/protocol/10.1007%2F978-1-4939-8935-5_22>

Melting point of 56-57C for general purposes

Filter wax before use

Wax must be kept at high temp during process

Need warm plate, cold plate, molds, forceps, forceps warmer, wax, wax dispenser, any additives (or embedding center)

Reservoir of paraffin is liquid

Metal molds should be warm

Warm forceps with commercial warmer, Bunsen burner, or alcohol lamp

Heat wax to 2C above melting point

Tissue block should not touch edges of mold

Forceps heated only until wax just melts on them, no hotter

Preparation of formalin-fixed paraffin-embedded tissue for immunohistochemistry  
<https://doi.org/10.1016/B978-0-12-420067-8.00015-5>

Cut tissue into 3-4mm thickness, fix slices 48h. (better fixation) Store in PBS until processed.

Dehydrate in cassettes in 70% ethanol for 20min, then 2x 95%, 2x 100%

Xylene for 20min 2x

Take tissue out of cassette and embed in liquid wax in mold for 30min. Fill rest of mold with wax to make block. Make sure tissue is aligned carefully.

Allow paraffin to cool.

Sectioning:

Slice in cryostat (recommend 5-10um), place slice on surface of 35-40C water bath, put slide under water under the slice and lift up to catch slice, put slide in 65C oven(not hot plate) for 10-20min until paraffin starts to melt.

Abcam protocol  
<https://www.abcam.com/protocols/ihc-tissue-processing-protocol>

Book: Theory and Practice of Histological Techniques, Bancroft

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8:55am – put frozen nodose sample in -20C freezer

9:30am – put in PBS on ice